

Epigenetics application in the diagnosis and treatment of bladder cancer

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Introduction: Bladder cancer is the sixth most common cancer in the Western world. Patients with bladder cancer require close monitoring, which may include frequent cystoscopy and urine cytology. Such monitoring results in significant health care cost. The application of epigenetics may allow for a risk adapted approach and more cost-effective method of monitoring. A number of epigenetic changes have been described for many cancer sites, including the urinary bladder. In this review, we discuss the use of epigenetics in bladder cancer and the potential diagnostic and therapeutic applications.

Materials and methods: A comprehensive search of the English medical literature was conducted in PubMed using the terms microRNA regulation, DNA methylation, histone modification and bladder cancer.

Results: The most important epigenetic changes include DNA methylation, histone modification and microRNA regulation. Both DNA hypomethylation and hypermethylation have been associated with higher rate of cancer. The association of epigenetic changes with bladder cancer has led to the research of its diagnostic and prognostic implications as well as to the development of novel drugs to target these changes with the aim of achieving a survival benefit.

Conclusions: Recently, epigenetics has been shown to play a much greater role than previously anticipated in the initiation and propagation of many tumors. The use of epigenetics for the diagnosis and treatment of bladder cancer is an evolving and promising field. The possibility of reversing epigenetic changes may facilitate additional cancer treatment options in the future.

Key Words: histone acetylation, bladder cancer, DNA methylation, microRNA regulation

Introduction

Bladder cancer is the sixth most common cancer in the Western world.¹ In the United States alone, it is estimated that 74,690 new cases were diagnosed in 2014 with 15,580 projected disease-specific deaths.^{2,3}

The 5-year survival rate of bladder cancer has remained relatively low despite the development of various surgical and chemotherapeutic methods, mostly due to high rates of recurrence and metastatic spread.⁴ The development of novel therapeutic strategies through the identification of key genes

involved in the pathogenesis of bladder cancer has therefore become paramount.⁵ In addition, bladder cancer patients typically require frequent surveillance with cystoscopies, imaging and urine cytologies, which results in a high annual expenditure.⁶ Epigenetics may offer a potentially more cost-effective method of monitoring such patients. Epigenetics refers to inherited modifications that are not encoded in the DNA structure itself.⁷ Recently, it has been shown that epigenetics plays a larger than anticipated role in cancer biology among other cell processes. A number of epigenetic changes have been found in several cancers including colon, lung, breast, prostate, and urinary bladder.⁸ These changes to the genome present a possible therapeutic target due to their potential reversibility.⁹ In this review, we discuss the use of epigenetics in the diagnosis of bladder cancer and potential prognostic and therapeutic applications.

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Epigenetics

Epigenetics is the study of the changes in gene activity that are not caused by changes in the DNA structure. Epigenetics includes, but is not limited to DNA methylation, microRNA (*miRNA*) regulation and histone modification.¹⁰ DNA methylation involves the covalent addition of a methyl group from S-adenosyl-methionine to a nucleotide, mostly cytosine within *CpG* dinucleotides in the DNA.¹¹ This process is facilitated by DNA methyltransferases (*DNMT*). Post-transcriptional regulation of gene expression occurs through an epigenetic mechanism called RNA interference, which is mediated by a class of small noncoding RNAs called *miRNA*. De-regulation of *miRNA* expression resulting from epigenetic modifications occurring in transformed cells may lead to tumorigenesis.¹² Histone modification includes methylation, acetylation, and phosphorylation of the histone proteins in the chromatin material.

DNA methylation, hypermethylation and hypomethylation

Among epigenetic alterations, DNA methylation has been studied extensively.¹⁰ DNA methylation is typically the formation of a covalent bond of a methyl group to the 5 carbon position of a cytosine ring, although it can occur to adenine or thymine rings as well.⁸ DNA methylation often occurs in a *CpG* dinucleotide (*CpG* island), a dinucleotide in which a cytosine is directly followed by a guanine. Methylation of *CpG* islands has been associated with the silencing of tumor-suppression genes.¹³ This silencing can be transferred through several generations of cells. In methylation of *CpG* islands, both strands are methylated at the cytosine base. After replication, one strand is methylated due to semi-conservation. This state is known as hemimethylation. DNA methyltransferases have a high affinity for hemimethylated substrates, which methylate the remaining cytosine. In this way, the silencing persists through several cell divisions.

The hypermethylation of DNA has been established as an epigenetic abnormality observed in several malignant tumors, particularly in *CpG* islands.¹³ Therefore, analysis of hypermethylated *CpG* islands is a promising method of detection and classification of cancers.¹⁴ Hypermethylation involves silencing genes that perform tumor suppression, DNA repair and other key processes. Hypermethylation requires a number of enzymes called DNA methyltransferases (*DNMT*),¹⁵ Figure 1. Four members of the *DNMT* family have been identified in mammals: *DNMT1*, *DNMT3A*, *DNMT3B* and *DNMT3L*. *DNMT1* is the most essential for cancer cell growth.¹¹

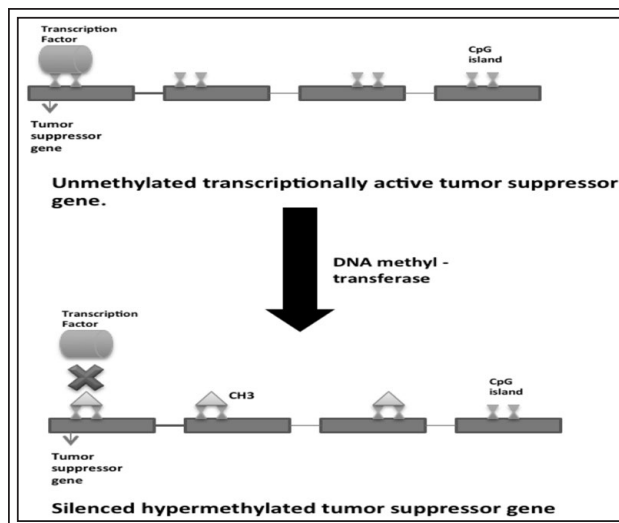


Figure 1. Epigenetic silencing of a tumor suppressor gene carried out by DNA methyl-transferase through hypermethylation of *CpG* dinucleotides.

DNA hypomethylation involves the removal of methyl groups from methylated DNA strands. It is facilitated by a group of enzymes known as demethylases.¹⁵ Similar to hypermethylation, DNA hypomethylation has also been associated with a higher risk of cancer.¹⁶ DNA hypomethylation is also a potential biomarker for bladder cancer. The gene *LINE-1* is one such marker.¹⁷ However, the relationship between this epigenetic alteration and bladder cancer is still unclear, and DNA hypomethylation has predominantly shown to be useful as a post-diagnosis marker.

MicroRNA regulation

Regulation of post-transcriptional gene expression takes place through the epigenetic mechanism of RNA interference. *MiRNAs* are a class of small noncoding RNAs that regulate this process. Within tumor cells, dysregulation of *miRNA* expression can occur, affecting its differentiation, proliferation and apoptosis. The overexpression of *miRNA* in tumor cells has been termed oncogenic whereas *miRNA* that shows reduced expression is referred to as a tumor suppressor.¹⁸ In a recent paper that assessed the expression of *miR-200c*, *miR-30b* and *miR-141* in tissue samples of patients with bladder cancer and healthy adjacent tissue, it was reported that *miR-141* was up-regulated in 91% of malignant tissue samples whereas *miR-200c* and *miR-30b* were over-expressed in 79% and 64% respectively.¹⁹ Yu et al recently reported the anti-metastatic properties of *miR-34a* by inhibiting epithelial mesenchymal transition (EMT) related proteins in bladder cancer. EMT is a transient process in which epithelial cells acquire a mesenchymal phenotype by

losing intercellular adhesions normally maintained by a group of proteins called *E-cadherins* with the subsequent increase in cell motility. Proteins related to EMT include *N-cadherin*, *Beta-catenin* and *Vimentin*. *MiR-34a* consistently promoted *E-cadherin* protein expression and inhibited *N-cadherin*, *Beta-catenin* and *Vimentin* protein expression. This data indicated that *miR-34a* functions as a tumor suppressor in bladder cancer.²⁰

Histone modification

Histones are proteins around which the double stranded DNA is coiled forming a structure known as nucleosome. Each nucleosome contains eight histone molecules (octamer): one pair each of *H2A*, *H2B*, *H3*, and *H4*. Both the histones and the DNA can be modified to silence or activate genes. Consequently, the proteins translated (or not translated) can cause the silencing or activation of other genes, creating a chain effect. Histone modification includes methylation, acetylation, and/or phosphorylation. Histone structure consists of two domains, a globular domain and a "tail" that contains the NH₂ terminal. Modification typically occurs on the "tail" domain at different amino acid positions. Nucleosomes are considered the basic unit of eukaryotic chromatin. There are essentially two types of chromatin: heterochromatin and euchromatin. Heterochromatin is highly condensed and is thus harder to transcribe, effectively silencing most genes within it. Euchromatin is more loosely packed and the genes are more easily transcribed. Like DNA methylation, identifying histone modifications has the potential for diagnosis or prognostication of bladder cancer. A well-studied histone modification is the trimethylation (*m3*) of histone 3 lysine 27 (*H3K27m3*). One study was able to correctly estimate gene expression from this marker in 65% of the subjects

studied.²¹ A list of genes involved in bladder cancer with its corresponding frequency is shown in Table 1.

Marker potential

Diagnosis

The use of epigenetics for the diagnosis of bladder cancer has great potential. It could reduce the need of invasive tests for tumor detection, including cystoscopy, which can miss some small or in-situ tumors despite good technique. DNA tests can be done easily using the polymerase chain reaction (PCR), which rapidly and accurately amplifies the subject's DNA, allowing it to be screened for genetic abnormalities.²² One of the most validated techniques to establish the presence or absence of methylation in a gene within a tumor sample is methylation-specific PCR (MSP). This technique requires only small quantities of DNA and allows for a rapid analysis of multiple markers.²³ DNA tests can be done by simply using voided urine from bladder cancer patients. A number of recent studies have found several potential diagnostic markers. One study found the genes *IRF8*, *p14* or *sFRP1* predicted the presence of bladder cancer with a sensitivity of 86.7% and a specificity of 94.7%. Methylation of *SFRP1*, *IRF8*, *APC* and *RASSF1A* were also indicative of a higher tumor grade.²⁴ Chung et al reported two sets of parallel genes, the first of which contained: *MYO3A*, *CA10*, *NKX6-2* and *DBC1* or *SOX11*, which had 81% sensitivity and 97% specificity for the detection of bladder cancer. The second set contained: *MYO3A*, *CA10*, *NKX6-2*, *DBC1* and *SOX11* or *PENK*, which had 85% sensitivity and 95% specificity. Thus, a total of six methylation markers were identified.²⁵ Yu et al studied an 11-gene set which confirmed bladder cancer with a sensitivity of 91.7% and an accuracy of 87%.²⁶ There are a number of genes that could be used as potential DNA markers including *APC*, *RASSF1A*, *SFRP1*, *GDF15*, *HSPA2*, *TMEFF2*, *HOXB2*, *KRT13*, and *FRZB*.^{6,14,27} The functions of these genes have been listed in Table 2.

Prognosis

A recent study has shown that epigenetic screening can be helpful in determining the prognosis of patients with bladder cancer. Methylation of the metastasis suppressor gene, *KISS1*, was shown to be associated with higher grade and higher stage tumors.²⁸ The *KISS1* gene has been identified as suppressing metastasis in melanoma and breast cancer cells. This gene encodes for a 145-amino acid protein that is processed into kisspeptins, which play a role at inhibiting cancer metastasis of different tumor types. In their report, Cebrian et al evaluated the effect of *KISS1* methylation on its expression and the clinical relevance in bladder cancer. Their results revealed that

TABLE 1. Mutated genes and involvement frequency in bladder cancer³⁷

| Gene | Frequency |
|--------|----------------------------|
| KDM6A | 20%-25% of bladder cancers |
| MLL2 | 27% of bladder cancers |
| ARID1A | 25% of bladder cancers |
| EP300 | 15% of bladder cancers |
| MLL | > 10% of bladder cancers |
| MLL3 | > 10% of bladder cancers |
| CREBBP | > 10% of bladder cancers |
| CHD7 | > 10% of bladder cancers |
| SRCAP | > 10% of bladder cancers |

TABLE 2. Genes and functions³⁸⁻⁴⁵

| Gene | Function |
|---------|--|
| APC | Cell proliferation, differentiation, migration, polarity |
| RASSF1A | Cell cycle regulation, apoptosis, microtubule stability |
| SFRP1 | Cellular senescence |
| GDF15 | Proliferation regulation and apoptosis |
| HSPA2 | Sperm maturity, function and fertility |
| TMEFF2 | Believed to be a tumor suppressor |
| HOXB2 | Decreases tumor proliferation |
| KRT13 | Encodes keratin |
| FRZB | Expressed in developing and mature chondrocytes |

KISS1 was hypermethylated in bladder cancer and that the methylation of the gene and its transcript expression could be used as a biomarker and a predictor of outcome for bladder cancer patients.²⁸

Therapeutic targets

Tumor suppressor gene silencing through an epigenetic mechanism is carried out through systematic enzyme catalyzed processes. Reactivation of these genes has become an attractive target for cancer therapy. There are several drugs that are being evaluated. These are broadly grouped into two categories: DNA methyl transferases (*DNMT*) inhibitors and histone deacetylase (*HDAC*) inhibitors.¹⁵

DNMT inhibitors

Nucleoside analogs are a large part of *DNMT* inhibitors. As mentioned previously, methylation typically occurs in CpG islands at the two cytosine bases (one on either strand).⁸ The understanding of the mechanism of methylation by *DNMT* suggests that inhibition of methylation can occur at a number of steps including recognition of the target sequence, flipping of the target base, formation of the initial covalent bond, methyl transfer to carbon 5 of cytosine and release of the substrate. Nucleoside analogs substitute the nitrogenous bases in DNA with analog structures to prevent methylation. These nucleosides are converted to deoxyribonucleoside triphosphates and subsequently incorporated into newly synthesized DNA in place of cytosine residues. *DNMT* is inactivated through binding to these newly synthesized residues that replace cytosine residues next to guanine, resulting in global hypomethylation.^{29,30} Zebularine is

one such drug, which is an analog of cytosine. Studies have shown that synthetic oligonucleotides containing Zebularine form tight complexes with bacterial methyltransferases leading to a potent inhibition of DNA methylation. One study tested whether Zebularine was an effective inhibitor of mammalian DNA methylation in human bladder carcinoma cells lines. It was noted that Zebularine could induce expression of a human tumor suppressor gene that had been silenced by methylation after just 48 hours of treatment.³¹⁻³³

HDAC inhibitors

There are a number of drugs that show *HDAC* inhibitory capacity. Due to their diversity, they are broadly categorized into five groups based on chemical composition and structure: short-chain fatty acids, hydroxamic acids, cyclic tetrapeptides, benzamides and aliphatic acids. The difficulty in using them lies in minimizing the toxicity of the drug, while maximizing the effect. Several *HDAC* inhibitory drugs have been tested for treatment of urological cancers including: Romidepsin, Vorinostat, and Panobinostat, however, trials have shown poor response and high toxicity levels.³⁴ Some reviews have suggested *HDAC* inhibitors show promise due to good bioavailability and, used in combination with cytotoxic chemotherapy, have acceptable toxicity. However their tumor inhibitory effect could not be independently assessed.³⁵ A combination of *HDAC* inhibitors and *DNMT* inhibitors can have a synergistic effect on apoptosis, differentiation and/or cell growth arrest in cancer cells.³⁶

Conclusions

Epigenetics has been shown to play a much greater role than previously thought in the initiation and propagation of tumors. The use of epigenetics for the diagnosis and treatment of bladder cancer appears to be a promising field. Drugs using epigenetic targets are undergoing tests for their toxicity, safety, and potential synergistic effects with other drugs. Epigenetic profiling also has the potential to identify patients at high risk of bladder cancer through non-invasive urine tests. Such testing could help identify patients at an earlier stage of the disease. The possibility of epigenetic changes being “reversed” makes such epigenetic modification therapy a powerful potential option for cancer treatment in the future. □

References

1. Negraes PD, Favaro FP, Camargo JL et al. DNA methylation patterns in bladder cancer and washing cell sediments: a perspective for tumor recurrence detection. *BMC Cancer* 2008;8:238.

2. Starke N, Singla N, Haddad A, Lotan Y. Long-term outcomes of high risk bladder cancer screening cohort. *BJU Int* 2015;April 18. Epub ahead of print.
3. Siegel R, Ma J, Zou Z, Jemal A. Cancer statistics, 2014. *CA Cancer J Clin* 2014;64(1):9-29.
4. van Lingen AV, Witjes JA. Current intravesical therapy for non-muscle invasive bladder cancer. *Expert Opin Biol Ther* 2013;13(10):1371-1385.
5. Jin Y, Lu J, Wen J, Shen Y, Wen X. Regulation of growth of human bladder cancer by miR-192. *Tumour Biol* 2015;36(5):3791-3797.
6. Marsit CJ, Houseman EA, Christensen BC et al. Identification of methylated genes associated with aggressive bladder cancer. *PLoS One* 2010;5(8):e12334.
7. Wayne TF. Epigenetics in the development, modification, and prevention of cardiovascular disease. *Mol Biol Rep* 2015;42(4):765-776.
8. Das PM, Singal R. DNA methylation and cancer. *J Clin Oncol* 2004;22(22):4632-4642.
9. Egger G, Liang G, Aparicio A, Jones PA. Epigenetics in human disease and prospects for epigenetic therapy. *Nature* 2004;429(6990):457-463.
10. Hoffman AM, Cairns P. Epigenetics of kidney cancer and bladder cancer. *Epigenomics* 2011;3(1):19-34.
11. Wang X, Zhang L, Ding N et al. Identification and characterization of DNazymes targeting DNA methyltransferase I for suppressing bladder cancer proliferation. *Biochem Biophys Res Commun* 2015;461(2):329-333.
12. Grange C, Collino F, Tapparo M, Camussi G. Oncogenic microRNAs and renal cell carcinoma. *Front Oncol* 2014;4:49.
13. Dudzic E, Miah S, Choudhry HM et al. Hypermethylation of CpG islands and shores around specific microRNAs and mirtrons is associated with the phenotype and presence of bladder cancer. *Clin Cancer Res* 2011;17(6):1287-1296.
14. Neuhausen A, Florl AL, Grimm MO, Schulz WA. DNA methylation alterations in urothelial carcinoma. *Cancer Biol Ther* 2006;5(8):993-1001.
15. Manoharan M, Ramachandran K, Soloway MS, Singal R. Epigenetic targets in the diagnosis and treatment of prostate cancer. *Int Braz J Urol* 2007;33(1):11-18.
16. Woo HD, Kim J. Global DNA hypomethylation in peripheral blood leukocytes as a biomarker for cancer risk: a meta-analysis. *PLoS One* 2012;7(4):e34615.
17. Patchesung M, Broonla C, Amnatrakul P et al. Long interspersed nuclear element-1 hypomethylation and oxidative stress: correlation and bladder cancer diagnostic potential. *PLoS One* 2012;7(5):e37009.
18. Gowrishankar B, Ibragimova I, Zhou Y et al. MicroRNA expression signatures of stage, grade, and progression in clear cell RCC. *Cancer Biol Ther* 2014;15(3):329-341.
19. Mahdavinzhad A, Mousavi-Bahar SH, Poorolajal J et al. Evaluation of miR-141, miR-200c, miR-30b Expression and Clinicopathological Features of Bladder Cancer. *Int J Mol Cell Med* 2015;4(1):32-39.
20. Yu G, Yao W, Xiao W, Li H, Xu H, Lang B. MicroRNA-34a functions as an anti-metastatic microRNA and suppresses angiogenesis in bladder cancer by directly targeting CD44. *J Exp Clin Cancer Res* 2014;33(1):779.
21. Dudzic E, Gogol-Doring A, Cookson V, Chen W, Catto J. Integrated epigenome profiling of repressive histone modifications, DNA methylation and gene expression in normal and malignant urothelial cells. *PLoS One* 2012;7(3):e32750.
22. Garibyan L, Avashia N. Polymerase chain reaction. *J Invest Dermatol* 2013;133(3):e6.
23. Andres G, Ashour N, Sanchez-Chapado M, Roper S, Angulo JC. The study of DNA methylation in urological cancer: present and future. *Actas Urol Esp* 2013;37(6):368-375.
24. Chen PC, Tsai MH, Yip SK et al. Distinct DNA methylation epigenotypes in bladder cancer from different Chinese sub-populations and its implication in cancer detection using voided urine. *BMC Med Genomics* 2011;4:45.
25. Chung W, Bondaruk J, Jelinek J et al. Detection of bladder cancer using novel DNA methylation biomarkers in urine sediments. *Cancer Epidemiol Biomarkers Prev* 2011;20(7):1483-1491.
26. Yu J, Zhu T, Wang Z et al. A novel set of DNA methylation markers in urine sediments for sensitive/specific detection of bladder cancer. *Clin Cancer Res* 2007;13(24):7296-7304.
27. Costa VL, Henrique R, Danielsen SA et al. Three epigenetic biomarkers, GDF15, TMEFF2, and VIM, accurately predict bladder cancer from DNA-based analyses of urine samples. *Clin Cancer Res* 2010;16(23):5842-5851.
28. Cebrian V, Fierro M, Orenes-Piñero E et al. KISS1 methylation and expression as tumor stratification biomarkers and clinical outcome prognosticators for bladder cancer patients. *Am J Pathol* 2011;179(2):540-546.
29. Sheikhejad G, Brank A, Christman JK et al. Mechanism of inhibition of DNA (cytosine C5)-methyltransferases by oligodeoxyribonucleotides containing 5,6-dihydro-5-azacytosine. *J Mol Biol* 1999;285(5):2021-2034.
30. Brank AS, Eritja R, Garcia RG et al. Inhibition of HhaI DNA (Cytosine-C5) methyltransferase by oligodeoxyribonucleotides containing 5-aza-2'-deoxycytidine: examination of the intertwined roles of co-factor, target, transition state structure and enzyme conformation. *J Mol Biol* 2002;323(1):53-67.
31. Cheng JC, Matsen CB, Gonzales FA et al. Inhibition of DNA methylation and reactivation of silenced genes by zebularine. *J Natl Cancer Inst* 2003;95(5):399-409.
32. Hurd PJ, Whitmarsch AJ, Baldwin GS et al. Mechanism-based inhibition of C5-cytosine DNA methyltransferases by 2-H pyrimidinone. *J Mol Biol* 1999;286(2):389-401.
33. Zhou L, Cheng X, Connolly BA et al. Zebularine: a novel DNA methylation inhibitor that forms a covalent complex with DNA methyltransferases. *J Mol Biol* 2002;321(4):591-599.
34. O'Rourke CJ, Knabben V, Bolton E et al. Manipulating the epigenome for the treatment of urological malignancies. *Pharmacol Ther* 2013;138(2):185-196.
35. Adam RM. Histone deacetylase inhibitors and bladder cancer. *J Urol* 2010;183(6):2120-2121.
36. Li LC, Carroll PR, Dahiya R. Epigenetic changes in prostate cancer: implication for diagnosis and treatment. *J Natl Cancer Inst* 2005;97(2):103-115.
37. Martin-Doyle W, Kwiatkowski DJ. Molecular biology of bladder cancer. *Hematol Oncol Clin North Am* 2015;29(2):191-203.
38. Andersen A, Jones DA. APC and DNA demethylation in cell fate specification and intestinal cancer. *Adv Exp Med Biol* 2013;754:167-177.
39. Fernandes MS, Carneiro F, Oliveira C, Seruca R. Colorectal cancer and RASSF family--a special emphasis on RASSF1A. *Int J Cancer* 2013;132(2):251-258.
40. Elzi DJ, Song M, Hakala K, Weintraub ST, Shio Y. Wnt antagonist SFRP1 functions as a secreted mediator of senescence. *Mol Cell Biol* 2012;32(21):4388-4399.
41. Zimmers TA, Jin X, Hsiao EC et al. Growth differentiation factor-15/macrophage inhibitory cytokine-1 induction after kidney and lung injury. *Shock* 2005;23(6):543-548.
42. Filipczak PT, Piglowski W, Glowala-Kosinska M, Krawczyk Z, Scieglińska D. HSPA2 overexpression protects V79 fibroblasts against bortezomib-induced apoptosis. *Biochem Cell Biol* 2012;90(2):224-231.
43. Lin K, Taylor JR Jr, Wu TD et al. TMEFF2 is a PDGF-AA binding protein with methylation-associated gene silencing in multiple cancer types including glioma. *PLoS One* 2011;6(4):e18608.
44. Boimel PJ, Cruz C, Segall JE. A functional in vivo screen for regulators of tumor progression identifies HOXB2 as a regulator of tumor growth in breast cancer. *Genomics* 2011;98(3):164-172.
45. Yamada A, Iwata T, Yamato M, Okano T, Izumi Y. Diverse functions of secreted frizzled-related proteins in the osteoblastogenesis of human multipotent mesenchymal stromal cells. *Biomaterials* 2013;34(13):3270-3278.